

PHOTOCHEMICAL SYNTHESIS
OF DEUTERIUM LABELLED 4-N-SUBSTITUTED CYTOSINES

Lech Celewicz, Jarosław Spychała and Krzysztof Golankiewicz*
Adam Mickiewicz University, Faculty of Chemistry, Grunwaldzka 6
60-780 Poznań, Poland

Summary

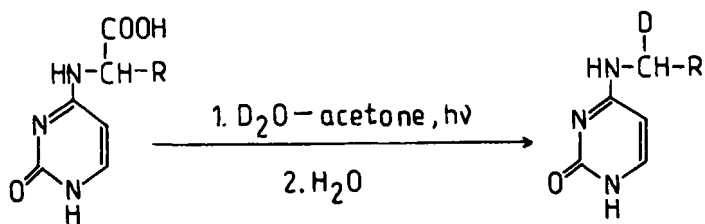
The efficient photochemical method for the preparation of 4-N-(1-deuterio-alkyl)cytosines using as precursors 4-N-(1-carboxy-alkyl)cytosines is described.

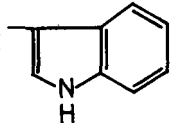
Key words: 4-N-(1-deuterio-alkyl)cytosines, photodecarboxylation.

Cytosine derivatives have received considerable attention due to antiviral and anticancer properties of the corresponding nucleosides^{1,2}. Among the large number of known cytosine analogs, 4-N-substituted derivatives are of interest as naturally occurring constituents of t-RNA³. We have recently reported that 4-N-(1-carboxy-alkyl)cytosines undergo remarkably efficient photochemical decarboxylation to give the proper 4-N-substituted cytosines⁴.

The present paper describes photochemical synthesis of 4-N-(1-deuterio-alkyl)cytosines. Ultraviolet irradiation with wavelengths longer than 290 nm in deuterium oxide-acetone (1:1, v/v) solution of compounds 1a-f^{4,5} and methanol-d-acetone (3:7, v/v) solution of 1g⁵ resulted in formation of photoproducts 2a-g containing a deuterium at the position formerly occupied by the carboxyl.

* To whom correspondence should be addressed.



- 1a, R=H
 1b, R=CH₃
 1c, R=CH(CH₃)CH₃
 1d, R=CH₂OH
 1e, R=CH₂COOH
 1f, R=CH₂Ph
 1g, R=CH₂-

	Yield(%)
2a	85
2b	79
2c	70
2d	45
2e	61
2f	67
2g	66

The irradiation of 4-N-(1-carboxy-2-hydroxyethyl)cytosine 1d has to be carefully controlled by tlc chromatography because initially produced 2d under prolonged irradiation undergoes further photochemical reaction to give 4-N-methylcytosine. However it was possible to obtain 2d in 45% yield. In case of photolysis of 4-N-(1,2-dicarboxy-ethyl)cytosine 1e only carboxyl group at the 1 position is decarboxylated and 4-N-(1-deuterio-2-carboxy-ethyl)-cytosine 2e is photochemically stable.

Thus the photodecarboxylation of 4-N-(1-carboxy-alkyl)-cytosines in the presence of deuterium oxide offers an experimentally simple method to prepare 4-N-(1-deuterio-alkyl)-cytosines of high isotopic purity in good yield. The isotopic purity of obtained compounds 2a-g as determined by NMR spectroscopy was higher than 97%.

In conclusion, we have developed a highly efficient method for the synthesis of the specifically deuterated 4-N-substituted cytosines. The utility of the procedure was demonstrated by the synthesis of seven 4-N-(1-deuterio-alkyl)cytosines.

EXPERIMENTAL

Elemental analyses were made on an Elemental Analyser Perkin-Elmer 240. ¹H- and ¹³C-NMR spectra were determined on a Jeol FX 90 Q (90 MHz) and in case of 2g on a Varian (300 MHz) spectrometer. Mass spectra were made on a Jeol JMS-D-100 mass spectrometer. Irradiations at $\lambda > 290$ nm were carried out with a 400 W high pressure mercury lamp with a cylindrical Pyrex light filter 1.5 mm thick.

Compounds 1a-g were prepared according to the literature method⁵.

Synthesis of 4-N-(1-deuterio-alkyl)cytosines 2a-g

Dry 1 (250 mg) was dissolved in deuterium oxide and the solution was evaporated to dryness in vacuum. Then the residue was dissolved in deuterium oxide-acetone (1:1, 100 ml) and irradiated under dry nitrogen atmosphere for 15-50 min, until TLC showed disappearance of 1. After irradiation the solution was evaporated, dissolved in water and again evaporated to dryness. The residue was subjected to chromatography on a silica gel column which eluted with chloroform-methanol 7:3 (v/v) gave 2. Recrystallization from water-ethanol afforded analytically pure samples of the products.

In case of 1g instead of deuterium oxide methanol-d was used due to poor solubility of 1g in deuterium oxide.

2a:

¹H-NMR (DMSO-d₆) δ 2.74 (d, 2H, -CDH₂), 5.68 (d, 1H, C⁵-H), 7.30 (d, 1H, C⁶-H), 7.78 (t, 1H, N⁴-H), 10.40 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 26.39 (t, -CDH₂), 93.41 (C⁵), 140.98 (C⁶), 156.96 (C²), 164.98 (C⁴).

MS, m/z (rel. int.) 126 (100), 110 (11), 96 (15), 82 (21).

2b:

¹H-NMR (DMSO-d₆) δ 1.08 (d, 3H, CH₃), 2.97-3.24 (m, 1H, CDH), 5.64

(d, 1H, C⁵-H), 7.27 (d, 1H, C⁶-H), 7.72 (d, 1H, N⁴-H), 10.33 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 14.09 (CH₃), 34.08 (t, CDH), 93.79 (C⁵), 141.14 (C⁶), 157.39 (C²), 164.43 (C⁴).

MS, m/z (rel. int.) 140 (100), 125 (50), 111 (50), 95 (27).

2c:

¹H-NMR (DMSO-d₆) δ 0.88 (d, 6H, CH₃), 1.61-1.99 (m, 1H, CH), 3.04 (m, 1H, CDH), 5.66 (d, 1H, C⁵-H), 7.26 (d, 1H, C⁶-H), 7.62 (d, 1H, N⁴-H), 10.31 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 20.05 (two CH₃), 27.36 (CH), 46.70 (t, CDH), 93.46 (C⁵), 141.09 (C⁶), 156.85 (C²), 164.76 (C⁴).

MS, m/z (rel. int.) 168 (34), 153 (14), 125 (100), 111 (45).

2d:

¹H-NMR (DMSO-d₆) δ 3.36 (m, 1H, CDH), 3.53 (d, 2H, CH₂), 4.34 (br s 1H, OH), 5.77 (d, 1H, C⁵-H), 7.35 (d, 1H, C⁶-H), 7.89 (d 1H, N⁴-H), 10.50 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 42.23 (t, CDH), 59.55 (CH₂), 93.79 (C⁵), 141.25 (C⁶), 157.23 (C²), 164.76 (C⁴).

MS, m/z (rel. int.) 156 (3), 138 (95), 137 (100), 126 (10), 125 (22), 112 (55), 111 (45).

2e:

¹H-NMR (DMSO-d₆) δ 2.46 (d, 2H, CH₂), 3.37 (m, 1H, CDH), 5.60 (d, 1H, C⁵-H), 7.25 (d, 1H, C⁶-H), 7.64 (br s, 1H, N⁴-H), 10.24 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 33.32 (CH₂), 35.49 (t, CDH), 93.13 (C⁵), 141.30 (C⁶), 156.48 (C²), 164.49 (C⁴), 172.73 (C=O).

MS, m/z (rel. int.) 166 (9), 138 (5), 137 (5), 125 (3), 111 (38), 73 (100).

2f:

¹H-NMR (DMSO-d₆) δ 2.83 (d, 2H, CH₂), 3.53 (m, 1H, CDH), 5.75 (d, 1H, C⁵-H), 7.25-7.53 (m, 6H, C⁶-H and C₆H₅), 7.99 (br s, 1H, N⁴-H), 10.31 (br s, 1H, N¹-H).

¹³C-NMR (DMSO-d₆) δ 34.46 (CH₂), CDH signals are covered by DMSO-d₆, 93.68 (C⁵), 125.97 (C₆H₅), 128.11 (C₆H₅), 128.52 (C₆H₅), 139.35 (C₆H₅), 141.14 (C⁶), 157.01 (C²), 164.43 (C⁴).

MS, m/z (rel. int.) 216 (4), 125 (15), 112 (14), 105 (49), 91 (100).

2g:

¹H-NMR (CD₃-OD) δ 3.02 (d, 2H, CH₂), 3.67 (m, 1H, CDH), 5.73 (d, 1H, C⁵-H), 6.98 (dd, 1H, C^{5'}-H), 7.03-7.17 (m, 2H, C^{2'}-H and C^{6'}-H), 7.27 (d, 1H, C^{7'}-H), 7.32 (d, 1H, C^{4'}-H), 7.55 (d, 1H, C⁶-H).

¹³C-NMR (CD₃OD) δ 28.50 (CH₂), 44.85 (t, CDH), 99.23 (C⁵), 115.02 (C^{7'}), 116.02 (C^{8'}), 122.10 (C^{6'}), 122.95 (C^{4'}), 125.05 (C^{5'}), 126.23 (C^{2'}), 131.60 (C^{8'}), 140.92 (C^{6'}), 144.69 (C⁶), 163.41 (C²), 169.41 (C⁴).

MS, m/z (rel. int.) 255 (1), 145 (9), 144 (16), 130 (48), 126 (70), 111 (100).

ACKNOWLEDGEMENT

This work was supported by the Grant RP II 13.2.3.

REFERENCES

1. De Clerq E. - Pure Appl. Chem. 55: 623 (1983).
2. Robins R.K. - Chem. Eng. News 64: 28 (1986).
3. Hall R.H. - The Modified Nucleosides in Nucleic Acids, Columbia University Press, New York 1971, p. 295.
4. Celewicz L., Spychała J., Golankiewicz K. - Synth. Commun. (in the press).
5. Ueda T., Fox J.J. - J. Med. Chem. 6: 697 (1963).